

System Set-up and Maintenance

Teacher Instruction Checklist:

Please add these items to your calendar, where appropriate.

Before tank set-up and seed planting:

- ☐ Make one laminated copy of the Data Log for each growth chamber. Use a dry erase marker to collect data and then enter it online. Put the log somewhere prominent, where it will be easy to fill out on a regular basis.
- ☐ Find good location for growth chambers– they will be heavy once they are filled with water. For wild celery, set aside 24 cups of sand for seed dispersal later. Sand should be as dry as possible. Set it aside to dry out for several days if necessary.
- ☐ Plant wild celery seeds by March 1st.

After tank set-up and seed planting:

- ☐ Send grant application to Chesapeake Bay Trust for transportation costs for planting events.
- ☐ Check water level and height of lights above water every week.
- ☐ If seeds have not germinated 2 full weeks after planting, call Mark Lewandowski at (410) 260-8634. Look Closely!
- ☐ Enter data on-line at the DNR website at <http://mddnr.chesapeakebay.net/bgic/loginindex.cfm>. If you do not have access to the Internet, fax data to MD-DNR each week, 2 pages per week at (410) 260-8859.
- ☐ Make sure to fill growth chambers with water before spring break.

At end of project:

- ☐ Prepare grasses for transport to planting event. Follow instructions in this section.



Materials:

Total List for 2 growth chambers

(Note: All materials will be provided by Maryland DNR/CBF unless otherwise noted.)

- 2 - growth chambers
- 2 - sponge filters
- 2 - power heads
- 4 - incandescent light bulbs (75 watt)
- 4 - swing arm desk lamp
- 2 - power strips with surge protectors
- 2 - ground fault interrupters (GFI)
- 2 - thermometers
- 2 - submersible aquarium heaters (Second Nature Acura 1000- 150 watt)
- 1 - pH test kit
- 1 - nitrate test kit
- 6-8 planting trays (depending on plant species)
- 1 - foam sheet
- 1 - set of plants (wild celery seed package* or redhead grass, water stargrass, or sago pondweed propagations)
- 1 - quart size freezer Ziploc bag**
- 1 - bag of topsoil (lower organic content than potting soil)
- 1 - bag of all-purpose sand
- 1 - **ruler or yardstick for height measurement
- 1 - **cup measure
- 1 - **5 gallon bucket (for mixing sediment and adding water)
- 1 - **about 100 feet of string (depending on the height of your ceilings) - optional

** **NOTE:** It is **extremely** important to keep wild celery seeds refrigerated (not frozen) until planting*

***Not provided*



Setting Up Your Growth Chambers:

Note: Read all instructions before beginning

Part 1: Separate seeds (**wild celery only**).

At least one day before you want to plant the wild celery seeds, break each pod into several pieces. Squeeze out the seeds and gelatinous substance that encases them into a jar of cold water. The gelatin will break down in about 24 hours. Place this jar in the refrigerator until you are ready to plant the seeds.

Part 2: Assemble growth chambers.

It will take one hour to completely set up one chamber. You can break the class into two groups and do both chambers at the same time. Depending on your experiment, you will need to adjust your tank assembly. Please read the specific directions in the Experiment Section of the manual.

Materials:

- 2 - growth chambers
- 4 - desk lamps
- 4 - incandescent light bulbs
- 2 - power heads
- 2 - sponge filters
- 2 - power strips
- 2 - ground fault interrupters (GFCI)
- 2 - submersible aquarium heaters (Second Nature Acura 1000-150 watt)

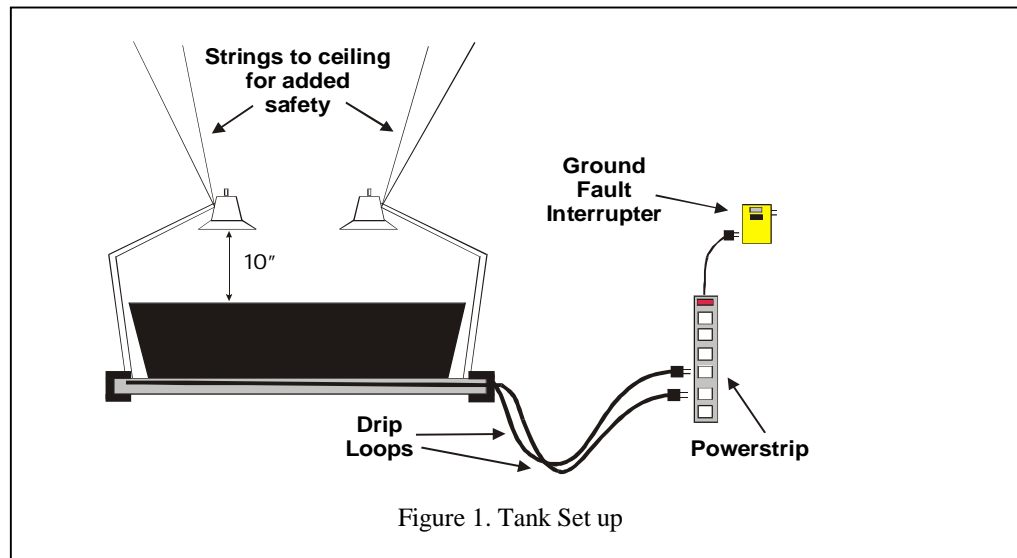
Procedure:

1. Once you have all the parts for your bay grass growth chamber, you will need to assemble them in your classroom. If split into two groups, a class of 15 students or more should be able to prepare the growth chamber in one hour. The growth chambers will be very heavy (approximately 100 lbs.) and difficult to move once filled, so choose your location carefully. If possible, do not place by a window.
2. Place chambers on table. Make sure the table has strong legs, and is near an empty wall outlet. Label the outside of each chamber clearly with an "A" or a "B".

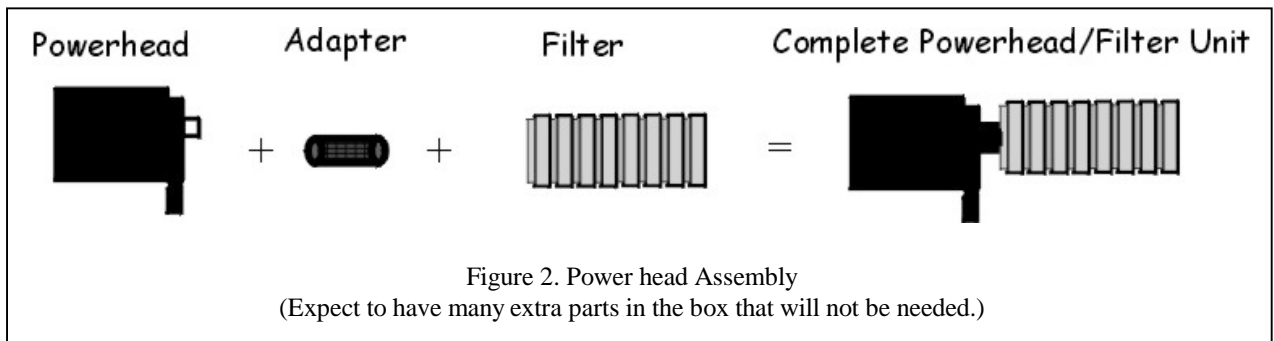


3. Assemble the lights and clamp them to the table so they can sit about 10 inches above the top of the chamber. Plug the lights into the power strip. Make sure a drip loop* is set up to prevent water from accidentally dripping into the power strip socket (see Figure 1).

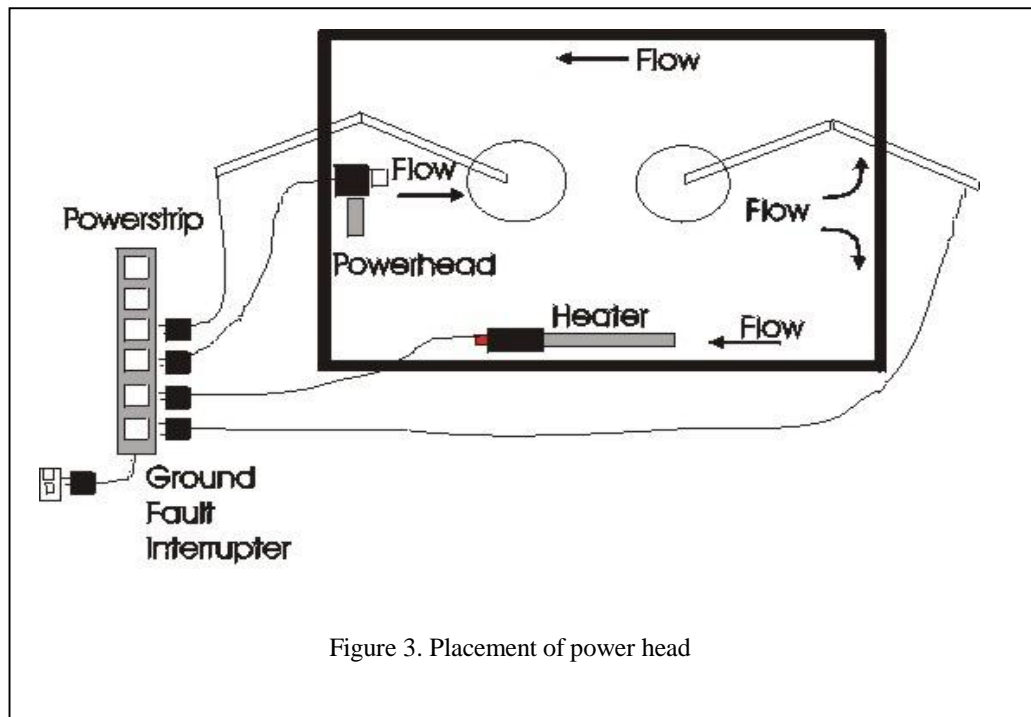
*It is up to the individual teacher to determine safety precautions to be taken with the lights. If accidentally submerged into the water, the light bulbs will burst and an electric shock of standard household current (120 volts) could result. Anything plugged into an outlet or power strip should have a drip loop to prevent water from accidentally dripping into the socket (see Figure 1).



4. Prepare the power head by attaching the cylindrical plug of the power head to the water intake of the power head. Then attach the sponge filter by stretching it over the adapter (see Figure 2). The sponge filter will prevent particles from clogging up the power head. In addition, it provides a medium for beneficial bacteria to grow. The bacteria will convert other forms of nitrogen to nitrate, which will help the plants to grow. DO NOT plug in the powerhead at this point.



5. Place the power head in the short end of the tank, directing the flow down the center of the tank (see Figure 3).



Part 3: Fill Growth Chambers.

Materials:

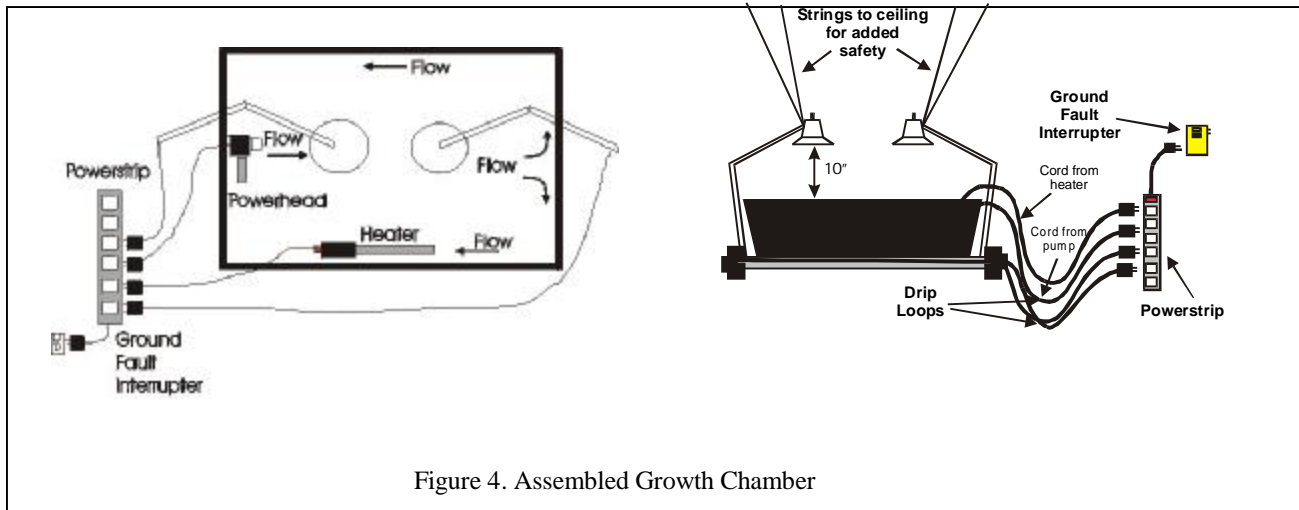
- 2 - bay grass growth chambers (from Step 2)
- 2 - thermometers
- 2 - submersible aquarium heaters
- water quality test kits

Procedure:

1. Fill each growth chamber with tap water to 10 cm (4 inches) deep.
2. Set the unplugged heater to the designated temperature:
 - Wild celery: both chambers at 75°F
 - Redhead grass/sago pondweed/water stargrass: chamber A: 75°F; chamber B: 84°F or 92°F
3. Lay the heater(s) in the bottom of the growth chamber. Plug heater(s) into the power strip. Set up drip loops on all cords so that water cannot run into the outlet (see Figure 4).
4. Plug the power strip into the GFCI and then into the outlet as in the diagram below.
5. Plug in the filter (and leave it on). It should immediately begin circulating water in your growth chambers. Remember to set up the drip loops!
6. Put the thermometer into the water. It can be attached to the side of each growth chamber with the suction cup, or it can float free.



7. The assembled growth chambers will look like this:



Preparing **Wild Celery** Planting Trays

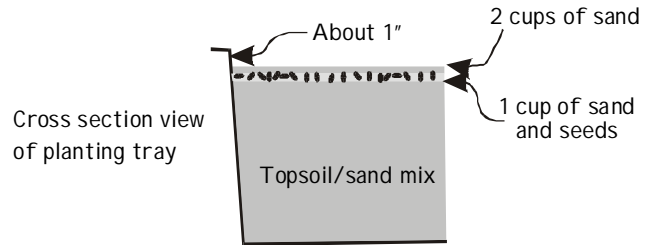
(Note: Read all instructions before beginning.)

It will take one hour to prepare all of the planting trays.

Materials:

- 1- container of bay grass seeds*
- 1- bag of sand
- 1- bag of topsoil
- 1- foam sheet
- 8- planting trays
- 1- quart-sized freezer Ziploc bag (Not provided)
- 1 - 5-gallon bucket (Not provided)
- 1 – Ruler or yardstick (Not provided)
- 1 - Cup measure (Not provided)

* It is extremely important to keep the wild celery seeds refrigerated (not frozen) until planting.



Procedure:

1. Set aside 24 cups of sand for use later when distributing seeds.
2. Thoroughly mix bag of topsoil and the remaining sand to a homogeneous mixture.
3. Use a permanent marker to label the lip of each of the 8 planting trays so that you can tell them apart (example: A1, A2, A3, A4, B1, B2, B3, B4).
4. Fill the 8 planting trays with the sediment mixture until all are equally full. Pack the sediment in each tray firmly with your fingertips.
5. Drain the seeds using filter paper or pour off the water using a spoon. Divide the seeds into 8 equal parts. The easiest way is to divide the pile in half, then each of those piles in half, etc.
6. In a quart-sized freezer Ziploc bag, add 1 of these 8 portions of seeds to 1 cup of the sand that was set aside. The sand should be as dry as possible before you add the seeds.
7. Shake this mixture in the quart size freezer Ziploc bag for 2 minutes.
8. Sprinkle this sand/seed mixture evenly over the sediment mixture that is in the planting tray.



9. Sprinkle an additional 2 cups of sand from the sand set aside previously over the sand/seed mixture. This should be a very thin layer (about 1/8 of an inch). If this layer is too thick, your seeds will not germinate.
10. Lay the foam sheet on top of the sediment surface. Remember, once the foam is in the water, it will float, so hold it in place tightly.
11. Two people should gently lower the trays into the first growth chamber. Tip one end of the tray when lowering, slowly. Hold the foam in place on top of the sediment surface until all bubbling has stopped. This may take quite a while (up to one minute). This will minimize disturbance of the sediment and movement of seeds.
12. Remove the foam sheet carefully by lifting one end slowly. Repeat steps 6 through 12 until each growth chamber has 4 trays in it.
13. Fill growth chambers with more water until water depth is 17 cm (7 inches). Check this level weekly and add water when necessary. Be careful not to disturb the sediment. Turn on the lights if not on already.
14. You are now ready to monitor your bay grass. See the next page for details. Remember: Week 1 = the first week that germination* is noticed.

*Wild celery seeds will typically take about 7 -10 days to germinate.



Preparing **Redhead Grass/Sago Pondweed/Water Stargrass** Planting Trays

(Note: Read all instructions before beginning.)

It will take one hour to prepare all of the planting trays.

Materials:

- 1 - Container of adult redhead grass/sago pondweed/water stargrass
- 1 - Bag of sand
- 1 - Bag of topsoil
- 6 - Planting trays
- 1 - Foam sheet
- 1 - 5 gallon bucket (Not provided)
- 1 - Cup measure (Not provided)

Procedure:

1. Set aside 12 cups of sand that will be used to cover the topsoil/sand mixture.
2. Thoroughly mix remaining sand and topsoil in a container (bucket, black tub, etc.).
3. Use a permanent marker to label the lip of each of the six planting trays or punch holes in the lip with a hole puncher so that you can tell them apart. Example: A1, A2, A3, B1, B2, B3.
4. Fill the 6 planting trays with the topsoil/sand mixture until all 6 are equally full to within about ¼ inch from the top. Pack the sediment in each tray firmly with your fingertips.
5. Sprinkle an additional 2 cups of sand that was set aside over the topsoil/sand mixture in each tray. This layer should be very thin. Spread sand evenly so the topsoil/sand mixture is no longer visible.
6. Lay the foam sheet on top of the sand/topsoil mixture surface. Remember, once the foam is in the water, it will float, so hold it in place tightly.
7. Two people should gently lower the trays into a growth chamber. Tip one end of the tray when lowering, slowly. Hold the foam in place on top of the sediment surface until all bubbling has stopped. This may take quite a while (up to one minute). This will minimize disturbance of the sediment and movement of plants.
8. Remove the foam sheet carefully by lifting one end slowly. Repeat steps 6 through 8 until each growth chamber has 3 trays in it.



9. Turn on lights, powerheads and make sure the heaters are set at the proper temperatures.
10. The parent tray will be placed in Chamber B. Fold all plants into the center of the tray, and gently cover them with the foam sheet. Lower the parent tray into the chamber as described in step 7.

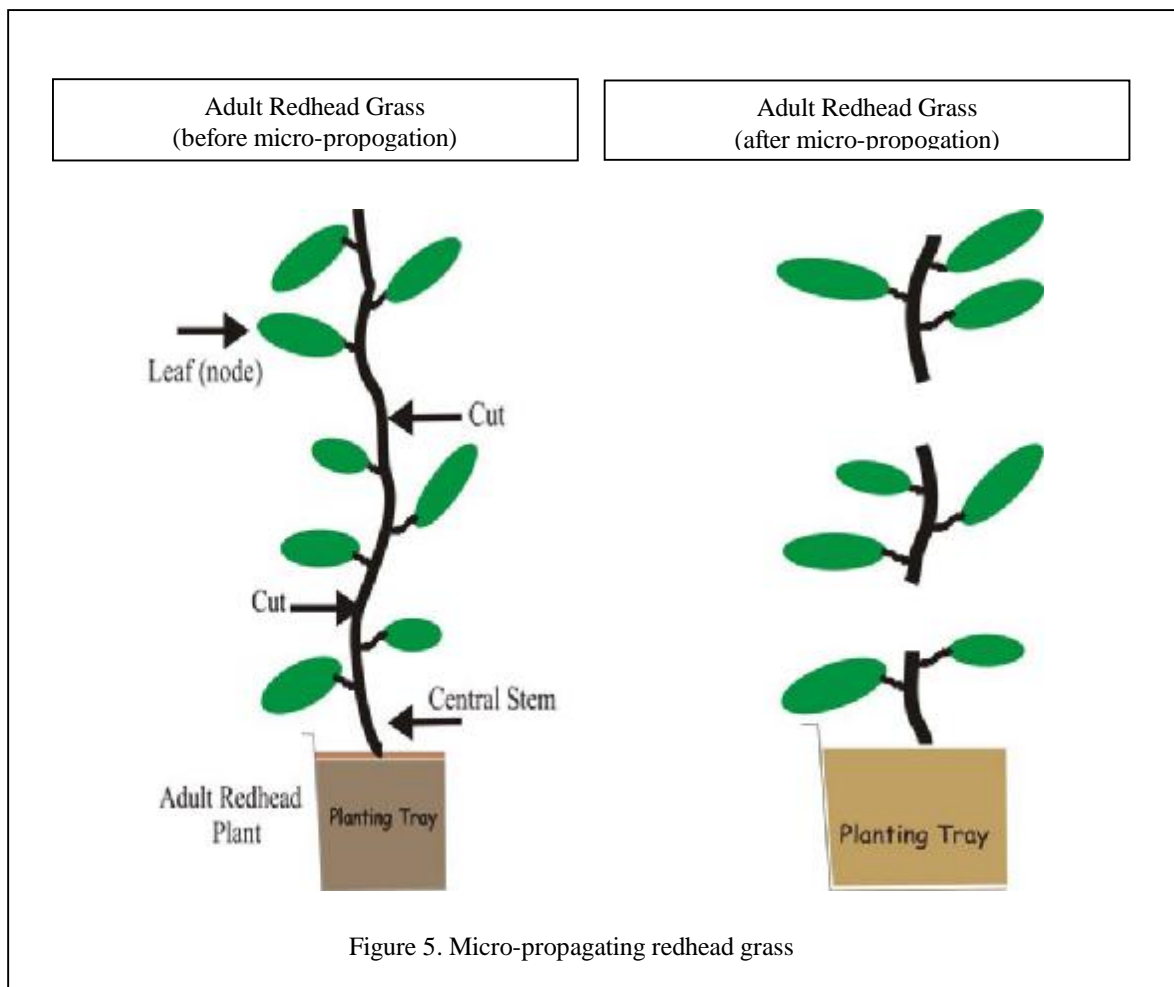


Micro-propagating **redhead grass/sago pondweed/water stargrass**

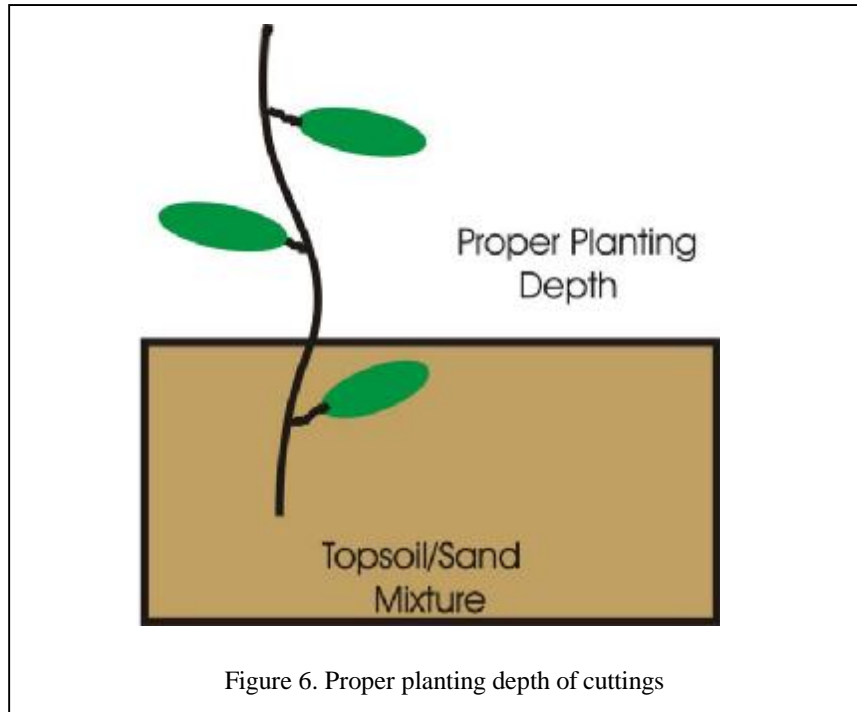
(Read all instructions before beginning. You will need an additional container of water to place the cuttings in during micro-propagation.)

It will take one hour to micro-propagate the plants from the parent tray.

1. When most of the plants in the parent tray have 8 leaves, it will be micro-propagation time. It will take several weeks for the plants to be large enough to micro-propagate. Lift each adult plant from the parent tray one at a time (without pulling the plants out of the sediment), holding them upright from the top of the central stem, and cut into segments of three leaves. Make sure to make the cut from the bottom and just above the leaf (node). Leave two leaves (nodes) on the rooted parent plant (Figure 5).
2. Put all of the cuttings into a container of water. The cuttings must remain wet at all times.



3. Divide the cuttings into two groups, one for each growth chamber. The number of cuttings will vary with each micro-propagation.
4. Plant all of the cuttings for growth chamber A into tray 1. Using the tip of your finger, bury each cutting so that one leaf is in the sediment and two leaves are above the sediment. Make sure they are evenly spaced from the sides of the tray and the other plants. Smooth the sand flat around the base of the stem (Figure 6).



5. Repeat until all of the cuttings for chamber A have been planted. Repeat the same procedure for chamber B.



The tray with the original adult plants will be put into growth chamber B (the elevated temperature growth chamber). There will only be three trays in growth chamber A. Your set-up should look like this after the first micro-propagation:

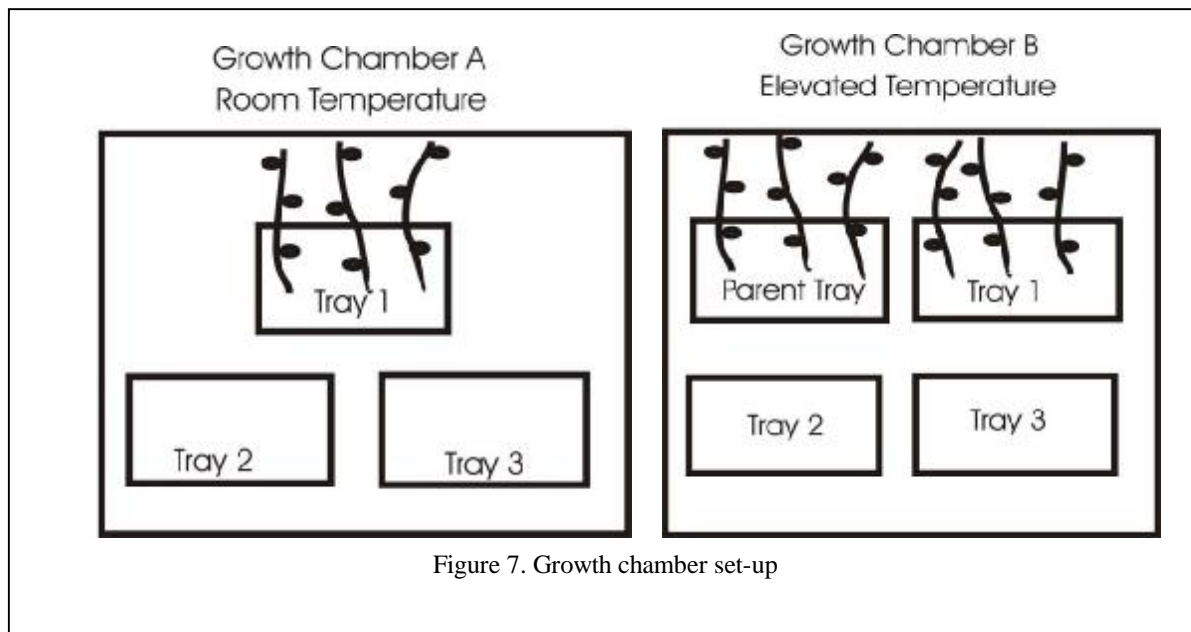


Figure 7. Growth chamber set-up

Check the water level weekly and add water when necessary. Be careful not to disturb the sediment. Trays two and three in both growth chambers will remain empty for several weeks. When a majority of the plants in tray one have eight leaves for both growth chambers, it will be time to do your second micro-propagation.

You will cut the plants in tray one of BOTH growth chambers into segments with at least three leaves (as explained in step 1). At the same time, micro-propagate the plants in the parent tray. Put all cuttings from both trays and the parent tray into the same container of water. You will have at least twice the amount of plants after micro-propagation.

Divide those cuttings into two groups, mixing the cuttings from each growth chamber. Following the directions above, plant the cuttings into tray two in each growth chamber. Tray three will remain empty until the 4 tallest plants in tray 2 of each growth chamber have grown eight leaves. When this occurs, the above process will be repeated again. Make sure you micro-propagate the parent tray at this time. After tray three has been planted, no more micro-propagation will be done.



Monitoring Your Bay Grass

Materials:

2 - thermometers

1 - ruler

Water chemistry test kits

“Bay Grasses in Classes: Data Log”

Procedure:

You will need to consistently monitor the growth of the bay grasses and the water quality in the growth chamber. A pair of students can easily do the daily monitoring during the first five minutes of each class period, and the weekly monitoring should take no longer than 20 minutes. Record your monitoring information in the “Bay Grasses in Classes: Data Log”.

1. Daily Monitoring (record the following information)

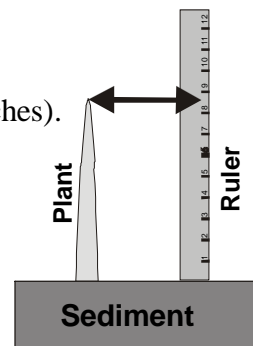
- Water temperature (in degrees Fahrenheit)
- Water level in the growth chamber. Fill to 16 ½ cm daily (or 6½ inches).
- Height of the light above the water surface. Should be 25 ½ cm (or 10 inches).
- Any additional observations or comments.

2. Weekly Monitoring

Wild Celery:

Week 1 = the first week that germination is noticed

- Water chemistry (pH, Nitrate). See instructions in box.
- Plant height above sediment (measured in cm, NOT inches).
 - Tallest plant in each tray and the average within each growth chamber.
- Any additional observations or comments.



Redhead Grass, Sago Pondweed, Water Stargrass:

- You will measure plant height and observe the number of leaves in tray one of both growth chambers (Figure 8). You will continue to send data from tray one until a majority of the plants have eight leaves (nodes). At this point, you will be ready to do your second micro-propagation.
- When you have divided your plants for the second time, you will only send data from tray two in each growth chamber. You will continue this until a majority of the plants in the tray have eight leaves, and you will micro-propagate for the third time. During each micro-propagation, make sure to clip the plants in the parent tray. After the third planting, all data sent will be from tray three.
- At this point, all of your trays will have plants in them. They will be roughly the same size. Continue to send data on tray three until planting day or MD-DNR contacts you and instructs you to stop recording.

3. Submitting your data

- Each week, go to the DNR on-line data entry page of the website at <http://mddnr.chesapeakebay.net/bgic/loginindex.cfm> to enter data for each growth chamber. If you do not have internet access, fax the completed Bay Grasses in Classes Data Log to Maryland DNR at 410-260-8859. Remember to send one sheet for each chamber each week. Your bay grass data will be compiled with data from all other participating schools and placed on the Bay Grasses in Classes website. You will be able to compare the growth of your plants with all the other participating schools by going to <http://mddnr.chesapeakebay.net/rastea/bgicsql/data.cfm>

Teachers Note: It is OK to ignore the plants for up to a week at a time **PROVIDED THE WATER LEVEL IS ADEQUATE**. If the water level drops below the powerhead intake, it will break, and an electrical shock could result. A water level drop of about 5 cm per week is normal, but this could vary significantly from school to school. Be sure that growth chambers are filled before leaving for spring break or long weekends



Preparing for Planting Your Bay Grass

Maryland DNR and CBF will identify a nearby site for bay grasses restoration. When your plants reach adult size (about 60 days) you will be able to assist scientists in planting your bay grasses at this restoration location. Your plants, along with those of the other participating schools, will increase the size of the new bay grass bed and will increase both its ecological value and its chance of survival.

To prepare your bay grass for transport to the planting site:

1. On the bay grass planting day, remove most of the water from the bay grass growth chamber by siphoning or dipping, until the water level just barely covers the top of the planting trays. This will reduce the weight, making the chambers easier to transport.
2. In order to reduce the drying of plants during transport, cover the growth chamber with several sheets of wet newspaper. These should be placed directly on the surface of the plants.
3. When you arrive at the planting location, be sure to put your grasses in an area out of direct sunlight and add water to the chambers if possible.
4. Make sure that you label the 2 growth chambers with your school's name and teacher's name, since there will be many other schools planting the same day.

Teacher's Note: Funding assistance for transportation to Bay Grasses in Classes planting locations will be provided by the Chesapeake Bay Trust. Please complete the attached application form and return to the Chesapeake Bay Trust ASAP to provide time for processing. If you do not submit the form before the due date, costs will not be covered.





2009 Wild Celery Data Log

School: _____

Teacher: _____

Grade/Class: _____

Week# 1 2 3 4 5 6 7 8 9 10 11 12 13 14
(week 1 = when germination is first noticed)

Experiment Type: (Circle one) Light Time, Flow Rate, Sediment Type, Temperature, Other

Chamber Type: _____ (for example, # of hours of light, flow, no flow, salinity tank)

Daily Monitoring

Date (month/day)	Water Temp (°F)	Water Depth (fill to 16 ½ cm)	Light Height (should be 25 ½ cm)	Comments (Date plants first visible, heavy algal growth)
Monday _____				
Tuesday _____				
Wednesday _____				
Thursday _____				
Friday _____				

Weekly Monitoring

Date	Average Temperature	pH	Nitrate (ppm)

Plant Height (centimeters)					
Growth Chamber A or B (indicate chamber type)					
Date	Tray 1	Tray 2	Tray 3	Tray 4	Average

***If you are doing a within tank experiment, please do not average the plant height data.**

NOTE: Please submit data at the end of each week via the online data entry system or by fax to Maryland DNR c/o Mark Lewandowski 410-260-8859.





System Set-Up and Maintenance
Bay Grasses in Classes



2009 Redhead Grass Data Log

School: _____ Teacher: _____

Grade/Class: _____ Week # _____ (Week 1 = first micro-propagation)

Chamber Type: (circle one) Chamber A Chamber B Chamber B
 75°F/24°C 84°F/29°C 92°F/33°C

Current week _____ Week of latest Micropropagation _____

Number of Micropropagations _____ When was tank treated with algicide _____
 (Black ink only please)

Daily Chamber Monitoring					
Date (month/day)	Water Temp (°F) (°C)		Water Depth (fill to 17 cm or 6 1/2")	Light Height (should be 26 cm or 10")	Comments (heavy algal growth, cloudy water)
Monday _____					
Tuesday _____					
Wednesday _____					
Thursday _____					
Friday _____					
Average Temp:					

Weekly Monitoring				
Date	pH		Nitrate	
Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Average Plant Height (cm)
Number of leaves	Number of leaves	Number of leaves	Number of leaves	Average number of leaves

NOTE: Please submit data at the end of each week via the online data entry system or by fax to Maryland DNR c/o Mark Lewandowski 410-260-8859.





System Set-Up and Maintenance
Bay Grasses in Classes



2009 Sago Pondweed Data Log

School: _____

Teacher: _____

Grade/Class: _____

Week # _____ (Week 1 = first micro-propagation)

Chamber Type: (circle one)

Chamber A
75°F/24°C

Chamber B
84°F/29°C

Chamber B
92°F/33°C

Current week _____ Week of latest Micropropagation _____

Number of Micropropagations _____ When was tank treated with algicide _____
(Black ink only please)

<i>Daily Chamber Monitoring</i>					
Date (month/day)	Water Temp (°F) (°C)		Water Depth (fill to 17 cm or 6 1/2")	Light Height (should be 26 cm or 10")	Comments (heavy algal growth, cloudy water)
Monday _____					
Tuesday _____					
Wednesday _____					
Thursday _____					
Friday _____					
Average Temp:					

Weekly Monitoring				
Date	pH		Nitrate	
Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Average Plant Height (cm)
Number of leaves	Number of leaves	Number of leaves	Number of leaves	Average number of leaves

NOTE: Please submit data at the end of each week via the online data entry system or by fax to Maryland DNR c/o Mark Lewandowski 410-260-8859.





System Set-Up and Maintenance
Bay Grasses in Classes



2009 Water Stargrass Data Log

School: _____

Teacher: _____

Grade/Class: _____

Week # _____ (Week 1 = first micro-propagation)

Chamber Type: (circle one)

Chamber A
75°F/24°C

Chamber B
84°F/29°C

Chamber B
92°F/33°C

Current week _____ Week of latest Micropropagation _____

Number of Micropropagations _____ When was tank treated with algicide _____
(Black ink only please)

<i>Daily Chamber Monitoring</i>					
Date (month/day)	Water Temp (°F) (°C)		Water Depth (fill to 17 cm or 6 1/2")	Light Height (should be 26 cm or 10")	Comments (heavy algal growth, cloudy water)
Monday _____					
Tuesday _____					
Wednesday _____					
Thursday _____					
Friday _____					
Average Temp:					

Weekly Monitoring				
Date	pH			Nitrate
Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Tallest plant in tray (cm)	Average Plant Height (cm)
Number of leaves	Number of leaves	Number of leaves	Number of leaves	Average number of leaves

NOTE: Please submit data at the end of each week via the online data entry system or by fax to Maryland DNR c/o Mark Lewandowski 410-260-8859.





System Set-Up and Maintenance
Bay Grasses in Classes



Consent/ Release Form

As parent or legal guardian of _____, I certify that said child has my permission to attend and participate in the "Bay Grasses in Classes" Program sponsored by the Maryland Department of Natural Resources. In signing this form, I acknowledge that my child will be participating in the following activities as part of this program: canoeing, seining, nature hikes, and planting grasses in Chesapeake Bay Tributaries. I also hereby grant the Maryland Department of Natural Resources the unconditional right to use the name, voice, and photographic likeness of _____ in connection with any of their audio video production, articles, or press releases, but not as an endorsement.

I, _____, hereby give my consent for Emergency Medical Care to be provided for my child, _____, while he/she is participating in the "Bay Grasses in Classes" program.

Physician: _____

Phone Number: (_____) _____

Allergies: _____

In case of an emergency during program hours, I can be reached at:

Signature of parent or legal guardian

Date

RELEASE STATEMENT:

I acknowledge that there are natural hazards with activities in an outdoor setting. I hereby affirm that my child is in good health and physically capable to perform the required activities of the program. In consideration of the Maryland Department of Natural Resources Bay Grasses in Classes Program accepting my child and to the extent permitted and approved by State Law, I hereby release and forever discharge the State of Maryland, its units, agents, and employees from all claims of liability for any damages or injuries which may be sustained while my child is at camp to the extent permitted by state law.

Parent/Guardian Signature

Date

Maryland Department of Natural Resources
Tidewater Ecosystem Assessment
Tawes State Office Building
580 Taylor Avenue D-2
Annapolis, MD 21401



*System Set-Up and Maintenance
Bay Grasses in Classes*



System Set-Up and Maintenance
Bay Grasses in Classes





Maryland Department of Natural Resources Resource Assessment Service Volunteer Information & Registration

This registration form between the MD Department of Natural Resources and each volunteer is subject to the following terms and conditions:

1. **Registration.** ALL volunteers must register on forms provided prior to doing any work. Registration qualifies the volunteers for State liability and medical protection.
2. **Duties.** No volunteer should undertake any work or use any equipment for which he/she is not trained and qualified. Volunteers are **not** permitted to drive a State Vehicle. Volunteers must use the same safety equipment required of the Department of Natural Resources personnel conducting similar activities. Volunteers who operate their own power equipment do so at their own risk.
3. **Liability.** Volunteers, like other State employees, are immune from tort liability if they are acting within the scope of their assigned public duties and without malice or gross negligence even if the damages exceed the limits of the State's waiver of immunity. Volunteers sued for alleged negligence are eligible for State legal assistance provided the limitations are not violated. The Department of Natural Resources relinquishes any claim for loss or damage to State property which results from supervised activities in State facilities, provided those activities are conducted without malice or gross negligence.
4. **Medical Coverage.** The State Treasurers Office provides volunteers with accident/medical insurance in the amount of \$2,500 for personal injuries and accidental death/dismemberment insurance in the amount of \$10,000. Proof of claim forms must be submitted to the Department of Natural Resources within thirty (30) days of the date of the accident.
5. **Coordination.** The volunteer program will be coordinated by Mark Lewandowski. Specific questions or concerns should be addressed to him at 410-260-8634. This form covers the period of _____5/4/09_____ to _____6/5/09_____.

Volunteer Name: _____

Address: _____

Telephone: _____

I acknowledge that I have read and understood the above information and understand that by signing this statement I am considered a DNR volunteer.

Signature _____

Date: _____





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Bay Grasses in Classes

